



# FORUM REVIEW ARTICLE

# Antioxidants in Translational Medicine

Harald H.H.W. Schmidt, Roland Stocker, Claudia Vollbracht, Gøran Paulsen, Dennis Riley, Andreas Daiber, and Antonio Cuadrado Antonio Cuadrado

#### **Abstract**

Significance: It is generally accepted that reactive oxygen species (ROS) scavenging molecules or antioxidants exert health-promoting effects and thus their consumption as food additives and nutraceuticals has been greatly encouraged. Antioxidants may be beneficial in situations of subclinical deficiency and increased demand or acutely upon high-dose infusion. However, to date, there is little clinical evidence for the long-term benefit of most antioxidants. Alarmingly, recent evidence points even to health risks, in particular for supplements of lipophilic antioxidants. Recent Advances: The biological impact of ROS depends not only on their quantities but also on their chemical nature, (sub)cellular and tissue location, and the rates of their formation and degradation. Moreover, ROS serve important physiological functions; thus, inappropriate removal of ROS may cause paradoxical reductive stress and thereby induce or promote disease. Critical Issues: Any recommendation on antioxidants must be based on solid clinical evidence and patient-relevant outcomes rather than surrogate parameters. Future Directions: Such evidence-based use may include site-directed application, time-limited high dosing, (functional) pharmacological repair of oxidized biomolecules, and triggers of endogenous antioxidant response systems. Ideally, these approaches need guidance by patient stratification through predictive biomarkers and possibly imaging modalities. Antioxid. Redox Signal. 23, 1130–1143.

# Introduction

Since the 1970s, oxidative stress has been evoked as a contributor to pathogenesis and thousands of studies have reported protective or therapeutic benefits of antioxidants in cellular and animal models of cardiovascular (43), neurodegenerative (62), and inflammatory diseases (75) and cancer (110) diseases. As a result, antioxidant supplements have been promoted as nutraceuticals and antioxidant vitamins, often with little or no clinical control or evidence.

# Failures and risks of antioxidants

However, with the exception of a few studies (50, 53, 67), antioxidants have almost always failed to show a significant effect in long-term clinical trials performed according to the criteria of evidence-based medicine (132). For example, the spin-trapping synthetic antioxidant NXY-059, developed for acute treatment of ischemia injury due to stroke, represents one of the most prominent and costly failures of a synthetic antioxidant ever clinically developed (135). Another example is the recent failure of the 2CARE study on the use of

Department of Pharmacology, CARIM, FHML, MIAS, Maastricht University, Maastricht, The Netherlands.

<sup>&</sup>lt;sup>2</sup>Victor Chang Cardiac Research Institute, Sydney, Australia.

<sup>&</sup>lt;sup>3</sup>University of New South Wales, Sydney, Australia.

<sup>&</sup>lt;sup>4</sup>Hochschule Fresenius, University of Applied Sciences, Idstein, Germany.

<sup>&</sup>lt;sup>5</sup>Norwegian School of Sport Sciences, Oslo, Norway.

<sup>&</sup>lt;sup>6</sup>Galera Therapeutics Inc., Malvern, Pennsylvania.

<sup>&</sup>lt;sup>7</sup>Labor für Molekulare Kardiologie, II. Medizinische Klinik und Poliklinik, Universitätsmedizin der Johannes Gutenberg-Universität, Mainz, Germany.

<sup>&</sup>lt;sup>8</sup>Centro de Investigación Biomédica en Red sobre Enfermedades Neurodegenerativas (CIBERNED), ISCIII, Madrid, Spain.

<sup>9</sup>Instituto de Investigaciones Biomédicas "Alberto Sols" UAM-CSIC, Madrid, Spain.

<sup>&</sup>lt;sup>10</sup>Instituto de Investigación Sanitaria La Paz (IdiPaz), Madrid, Spain.

Department of Biochemistry, Faculty of Medicine, Autonomous University of Madrid, Madrid, Spain.

<sup>\*</sup>These two senior authors contributed equally.

<sup>©</sup> Harald H.H.W. Schmidt *et al.* 2015; Published by Mary Ann Liebert, Inc. This Open Access article is distributed under the terms of the Creative Commons Attribution Noncommercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

coenzyme Q10 in the largest ever therapeutic trial for Huntington's disease (61b).

In some recent meta-analyses, a beneficial effect of antioxidants was claimed, such as for vitamin C in breast cancer (47), atrial fibrillation (2), stroke (25), or endothelial function (5). However, a more definitive proof of antioxidant benefit would have been required to measure plasma levels of the administered compounds. The latter was also a major limitation in several large clinical trials, for example, the Heart Outcomes Prevention Evaluation (HOPE) (137, 158) and the HOPE-The Ongoing Outcomes (17). Even worse than no effect, chronic use of multivitamins without clinical control, especially lipid-soluble antioxidants at dosages above the upper safety limit, may be associated with increased health risks (Table 1) (13).

In sedentary rats, vitamin E administration reduced liver oxidative damage, while in rats submitted to chronic exercise, vitamin E decreased antioxidant levels (148). It remains controversial whether chronic intake of high concentrations of certain antioxidants has a harmful effect on performance and whether this might be due to redox cycling reactions that could even convert an antioxidant into a pro-oxidant (18, 116, 125). Taken together, these observations indicate that to identify efficient redox-based therapeutic strategies, there is first a need to reevaluate the physiological and pathological relevance of reactive oxygen species (ROS), and then to determine whether an antioxidant approach is feasible and in which situations.

Another example of the failure of antioxidant therapy is provided in pre-eclampsia. Some studies have reported a modest increase in oxidative stress biomarker, F2-isoprostane, at late stages of pregnancy (112) as well as low levels of gamma-tocopherol may be considered as a risk factor for pre-eclampsia (63). However, concomitant

supplementation with vitamin C and vitamin E did not prevent pre-eclampsia in women at risk and, even worse, this treatment increased the rate of births with low weight (121).

Absorption, distribution, metabolism, and excretion of specific antioxidants are essential aspects of antioxidant therapy that have not been analyzed in detail for many compounds, especially for nutraceuticals. Yet, the pharmacokinetic (bioavailability and frequency of administration) and pharmacodynamic (therapeutic index and onset of action) properties of specific antioxidants are critical to assess their clinical usefulness (11). This is best exemplified with compounds that need to cross the blood–brain barrier (73).

### Redefining oxidative stress

The term, ROS, groups a number of oxygen-containing molecules with different chemical reactivity. ROS include not only  $O_2^{-\bullet}$ , hydroxyl radicals, alkoxyl and peroxyl radicals,  ${}^{\bullet}$ NO, and nitrogen dioxide radicals but also nonradical species such as hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), hypochlorite, peroxynitrite, singlet oxygen, lipid peroxides, and others. At the enzymatic level, the discovery of superoxide dismutases (SOD), catalase, peroxiredoxins, sulfiredoxins, and glutathione peroxidases implied that the formation of at least some forms of ROS, for example,  $O_2^{-\bullet}$ , can be harmful if not properly controlled (24, 90, 140).

At the chemical level, when talking about ROS and ROS scavengers, it is crucial to know what kinds of molecules are involved. For instance, in the case of ascorbate, this antioxidant avidly reacts with alkyl, peroxyl, and alphatocopheroxyl radicals, modestly with superoxide, and poorly with  $\rm H_2O_2$ . While ascorbate also reacts very rapidly with hydroxyl radicals, this unlikely contributes to its antioxidant

Table 1. Meta-Analyses and Reviews of Chronic Oral Vitamin and Antioxidant Substitution

Study type	Antioxidants assessed	Objectives and endpoint	Conclusions
Meta-analysis (150)	$\beta$ -Carotene and vitamin E	Effects on CVD-related and unrelated mortality and morbidity	β-Carotene possibly harmful; vitamin E no effect
Meta-analysis (14)	β-Carotene, vitamins A, C, E, and selenium, alone and mixed	Incidence of cancer and dose–response effect on mortality	High-dose vitamin E may increase mortality; selenium may be beneficial in the prevention of cancer; other antioxidants no effect
Review (81)	β-Carotene, vitamin E, and antioxidant cocktail	Primary and secondary prevention of CVD and progression of intima-to-media thickening	Data do not justify supplementing antioxidants beyond the current dietary guidelines
Meta-analysis (102)	Vitamin E alone and in combination	Dose–response effect on all-cause mortality rates	High-dose vitamin E may increase all-cause mortality and should be avoided.
Meta-analysis (133)	Vitamin E	All relevant CVD-related endpoints and lipid levels	Vitamin E does not affect CVD-related outcomes.
Meta-analysis (36)	Vitamin E alone and in combination	Odds ratio of relevant CVD-related endpoints	Vitamin E no effect
Meta-analysis (13)	β-Carotene, vitamins A, C, E, and selenium, alone and in combination	All-cause mortality with regard to other factors such as dose and type of supplement	$\beta$ -Carotene and vitamins A and E may increase mortality
Decision tree and Markov chain model (33)	Vitamin E alone and in combination	Event rates and quality- adjusted life year	Nonselective application of high-dose vitamin E does more harm than good

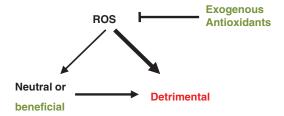


FIG. 1. Classical view on oxidative stress and antioxidants. Initially reactive oxygen species (ROS) were seen as a metabolic by-product that in low concentrations is neutral or beneficial (having signaling effects), but in higher concentrations can lead to a cellular, tissue, or even whole body redox imbalance with detrimental consequences. Hence, it was seen feasible to initiate systemic treatment with ROS scavengers as antioxidants to reverse this imbalance by reducing ROS levels. Such exogenous antioxidants were not expected to have any possible side effects. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

activity because hydroxyl radicals are so reactive that they react with essentially all molecules at diffusion-controlled rates so that ascorbate will always be outcompeted.

Originally, the term oxidative stress was primarily defined by quantitative deviation from a supposedly neutral steady-state level of ROS (136, 140). Quantitative deviation means a state where either increased formation of ROS or diminished cellular antioxidant defense leads to disequilibrium and a shift in the cellular redox balance (Fig. 1). However, we now know that for ROS, as for many other messengers, cellular levels are less relevant compared with subcellular concentrations (20, 68).

Quantity, that is, deviation from an overall equilibrium, on its own cannot explain or define oxidative stress. In fact, at least in many experimental models, only very moderate overall increases in ROS levels are observed, yet correlating with appreciable dysfunction and accumulation of relevant biomarkers (154). Therefore, in addition to quantitative accumulation of ROS, the oxidative stress concept needs to integrate other features of ROS such as their subcellular or tissue location, chemical nature (*e.g.*, O<sub>2</sub><sup>-•</sup>, H<sub>2</sub>O<sub>2</sub>, ONOO<sup>-</sup>), kinetics of formation and degradation, and time of exposure (Fig. 2) (76). Examples include the expression of NOX1 in hypertension, causing only a small increase in ROS (154), but *via* its caveolar localization, its product O<sub>2</sub><sup>-•</sup> interferes with \*NO formation and its vasodilator function (88); conversely, H<sub>2</sub>O<sub>2</sub> induces endothelial nitric oxide synthase (eNOS) and is a vasodilator (34, 100).

### Physiological functions of ROS

Another potential problem of antioxidants is the many physiological functions of ROS. The discovery of NOX isozymes, the only known enzyme family with the sole purpose of producing ROS, indicated that at least certain ROS are physiologically essential. For a long time, and still for newcomers to the field, it was considered that the increased formation of ROS equates to oxidative stress and represents a biochemical accident that can be prevented by systemic antioxidant treatment (mostly cell culture or animal-based studies using high doses of the drug). This misconception is based on the wrong assumption that ROS fulfill no physio-

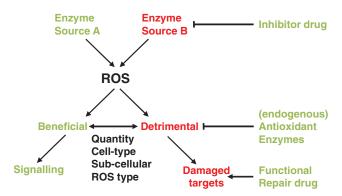


FIG. 2. Refined scheme and major changes in our understanding of oxidative stress and possible interventions. ROS are now viewed not necessarily as diseasetriggering molecules but also as essential signaling messengers. The distinction between beneficial and detrimental effects of ROS cannot be defined by overall cell, tissue, or whole body quantity of ROS formation. Instead, these differences may be due to different sources of ROS in different cellular and subcellular compartments (physiological and nonphysiological), the type of ROS being formed (e.g., superoxide vs. hydrogen peroxide). In light of the many negative results with broad systemic antioxidants, new therapeutic approaches may include more targeted or specific antioxidants, limiting antioxidants only to situations with acute and extremely high ROS levels, or indirect antioxidant mechanisms by upregulating endogenous antioxidant defense systems. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

logical function. The first description was the role of NOX2 (then termed gp91phox) in immunocyte oxidative burst.

Other NOX isoforms have different physiological roles in nonphagocytic cells, for example, in hearing (111), thyroid hormone synthesis (32) and angiogenesis, and the constriction of the ductus arteriosus (72). Furthermore, ROS play important roles in normal cellular function, including differentiation, proliferation, aging, and repair processes (124). Nitric oxide/nitrogen monoxide (\*NO) and H<sub>2</sub>O<sub>2</sub> seem to act primarily as messenger molecules, for example, they essentially contribute to vasodilation, proliferation (103), and promote or counteract programmed and spontaneous cell death (apoptosis and necrosis) (12, 147).

In working muscles, ROS are generated in the microcirculation, sarcolemma, and mitochondria (64) as part of a beneficial physiological response. Depending on mode, intensity, and duration of the exercise, antioxidants thus affect exercise capacity and adaptation processes (Fig. 3). For example, antioxidants, such as vitamins C and E, appear to blunt exercise-induced mitochondrial biogenesis (116); even anaerobic exercise, for example, strength training, can be affected with respect to muscle strength and hypertrophy (94, 117). Other observations indicate that vitamin C supplementation decreases oxidative stress and might increase exercise performance only in those subjects with low initial concentration of vitamin C (114).

Both the redefinition of oxidative stress (from a global redox disequilibrium to very subtle, even subcellular changes) and the many physiological functions of the same ROS that also cause disease may explain why systemic and chronically applied antioxidants may not necessarily have beneficial effects or even cause reductive stress or harm (9).

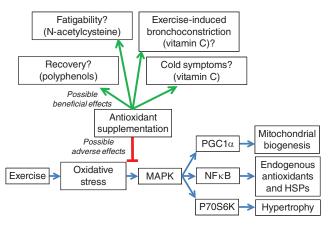


FIG. 3. Simplified scheme of antioxidant supplementation outcomes in exercising individuals. Possible adverse effects are linked to blunted cell signaling and adaptation to exercise by reducing oxidative stress. The adaptations will depend on the exercise mode (endurance vs. strength), but MAPKs may be important signaling proteins (65). The possible positive effects could not only be linked to the exercised muscles (reduced fatigability and improved recovery) but also the upper respiratory system by alleviating common cold symptoms and exercise-induced bronchoconstriction (10, 51, 52, 101). HSP, heat shock proteins (e.g., HSP70); JNK, c-Jun N-terminal kinases; MAPK, mitogen-activated protein kinases (p38, ERK1/2 and JNK); NFκB, nuclear factor kappa B (transcriptions factor); PGC1α, peroxisome proliferator-activated receptor gamma coactivator 1-alpha. Endogenous antioxidants include superoxide dismutase, catalase, and glutathione peroxidase. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

For example, ROS formed during the oxidative burst of phagocytic cells protect against infection and tumor cells, which imposes the risk of any chronically applied antioxidant (71, 118, 129). On the other hand, experimental data support the hypothesis that vitamin C affects epigenetic reprogramming toward less invasiveness of tumor cells (44, 149, 156).

# Paradoxes between in vitro versus in vivo and animal versus human studies

One potentially large problem in the field of ROS may be the use of *in vitro* models to observe cytoprotective effects of antioxidants. Standard cell culture, isolated organs, and cell-free models represent artifact-prone conditions due to a very high partial oxygen pressure of about  $0.2 \, \text{m}M$  (79). In addition, the frequently published exposure of cells to a bolus of high micromolar concentrations of an oxidant is nonphysiological, even if this may be reached in some subcellular compartments or intercellular spaces. It likely impairs cellular antioxidant defenses in a manner that is hardly ever seen *in vivo* (46).

Yet, it remains puzzling why so many *in vivo* animal studies that have shown beneficial effects of antioxidants were later not translatable into the clinic because they failed to show benefit or in some studies, HOPE (95, 157), HOPE-TOO (89), a prospective study with vitamin C in postmenopausal women with diabetes mellitus (82), and a prospective cohort study conducted among 83,639 healthy male US physicians (107), worsened outcome (Table 1). However, these and other studies often do not monitor plasma concentrations of the supplemented antioxidant. Measurement of drug levels in tissues where the dis-

ease process is to be modified by the drug is not always practical (e.g.), the central nervous system, heart) and target engagement biomarkers may be more helpful. Nevertheless, even in these cases, plasma levels of the drug serve important purposes, such as determination of drug safety and effective drug range.

Thus, it is possible that the lack of response had pharmacokinetic rather than pharmacodynamic reasons. In the European Prospective Investigation into Cancer (EPIC)-Norfolk study, vitamin C concentrations correlated with lower risk of all-cause mortality (77) and, in a meta-analysis of prospective studies, with a lower risk of stroke (25).

According to meta-analyses, long-term, high-dose N-acetylcysteine (NAC) treatment may improve the clinical complications of patients with chronic obstructive pulmonary disease (134) and the prophylactic use of NAC could reduce the incidence of postoperative atrial fibrillation and all-cause mortality in adult patients undergoing cardiac surgery (87). However, apart from a possible publication bias, such non-interventional observational studies and correlations cannot establish cause—effect relationships.

Other reasons for the failure of chronic oral antioxidant therapy may include the slow reaction constants between superoxide and vitamins C and E  $(3.3 \times 10^5 \text{ and } 4.9 \times 10^3 \text{ m}^{-1} \text{s}^{-1}$ , respectively, compared with  $1.9 \times 10^{10} \text{ mol}^{-1} \text{ s}^{-1}$  for the reaction constant between \*NO and superoxide) (141). Moreover, the inclusion criteria for patients can never be as restrictive as in preclinical studies, and in some cases, the disease may be at different stages of progression or clinical manifestation. In addition, as indicated before, in most cases, chronic, oral vitamin treatment has not been monitored consistently to provide plasma level concentrations or changes in target engagement biomarkers.

The differences between preclinical stages and clinical settings are also affected by differences in the antioxidant drug's mechanism of action and especially due to pharmacokinetic and pharmacodynamic constraints (11). This is best exemplified with compounds that need to cross the bloodbrain barrier (73).

Finally, it is interesting to note that basic research is biased by the lack of publications with negative results while most outcomes of clinical trials are reported. Therefore, it is possible that this paradox is due, in part, to an underrepresentation of the real outcomes in preclinical studies.

# Nonredox mechanisms of antioxidants

In other cases, the action of compounds with antioxidant activity may actually involve nonredox mechanisms, such as intercalation of DNA and/or inhibition of topoisomerase, DNA polymerase, and ribonucleotide reductase (40). Flavonoids and other phytochemicals can bind to functionally diverse cellular targets and this may modulate the activity of a large number of downstream genes (4).

#### **Ongoing Development of Direct Antioxidants**

Figure 4 depicts some conceptual strategies used to block the imbalanced production of ROS. Despite the failures and concerns with the use of antioxidant therapy, there are still several clinical studies and developments ongoing to translate the direct antioxidant principle into the clinic. Most of the antioxidant therapies so far utilized compounds that donate either one or two electrons, which require high

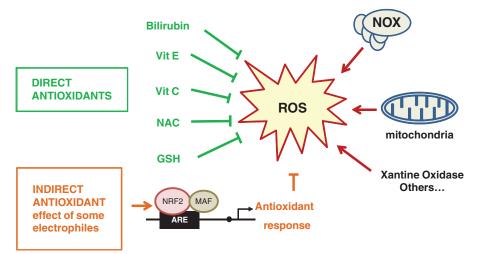


FIG. 4. Conceptual approach to the use of small molecules as direct and indirect antioxidants. The main sources of ROS production include NOX, mitochondria, xanthine oxidase, and other enzymes. Direct antioxidants (green) react with ROS and, if properly handled, may decrease oxidative stress. Indirect antioxidants (orange) activate cellular antioxidant defenses, such as NRF2mediated gene transcription, and provide a supplemental enzymatic armamentarium to maintain redox homeostasis. To see this illustration in color, the reader is referred to the web version of this article at www .liebertpub.com/ars

concentrations and impose the risk of converting the antioxidant into a pro-oxidant. To allow for lower doses, one alternative approach is to use compounds that are recycled by cellular reductases or act catalytically (90).

# Recycling antioxidants

Ascorbate typically reacts with free radicals, producing an ascorbyl radical intermediate that may be reduced back or undergo further oxidation to dehydroascorbic acid, for which specific reductases exist (86). Other recycling antioxidants include low-molecular-weight thiols (*e.g.*, glutathione and acetylcysteine), thioethers (*e.g.*, methionine), and many more. In the case of vitamin E ( $\alpha$ -tocopherol, in which case recycling depends on nonenzymatic reactions), BO-653 was developed as a synthetic, improved vitamin E-like molecule for the treatment of cardiovascular disease (intimal hyperplasia), but it failed in clinical trials (28). Other examples are the above-mentioned synthetic antioxidant NXY-059 that failed to improve outcomes in stroke patients (135) or coenzyme Q10, which failed to improve prognosis of patients with Huntington's disease (61b).

The glutathione peroxidase mimetic, ebselen (2-phenyl-1,2-benzoisoselenazol-3(2H)-one), was once considered a promising synthetic antioxidant since it reacts rapidly with peroxides (106, 153) and peroxynitrite (98), preventing lipid peroxidation and oxidative damage of other biomolecules. However, ebselen failed due to severe liver toxicity and inhibition of thiol-dependent enzymes (29). Compounds with a metal center such as SOD or catalase can in principle detoxify  $O_2^{-\bullet}$  and  $H_2O_2$  in a catalytic manner.

Trace elements such as zinc and selenium may augment antioxidant enzyme activity, but it will be difficult to ascribe their beneficial properties to modulating oxidative stress and not to other effects related to their function as cofactors of nonredox proteins. Metal porphyrins (FeTMPyP, FeTMPS, MnTBAP) were not only developed as SOD mimetics (61, 115) but unfortunately also showed pro-oxidative activities in biological systems and, so far, have not reached the stage of clinical application.

A possible alternative includes mitochondria-targeted SOD mimetics, for example, Mn(III) 5,10,15,20-tetrakis(N-methylpyridinium-2-yl)porphyrin (MnTM-2-PyP(5+)) (7, 8), or the so-called next-generation SOD mimetics. This compound class includes Mn-pentaazamacrocycles such as GC4403 (6,

128). This and other members of the Mn-pentaazamacrocyclic ligand class of SOD mimetics function by a one-electron redox cycle, with oxidation of Mn(II) being the rate-determining step (6).

In animal models of oxidative stress caused by total body irradiation, GC4403 increased survival, decreased intestinal apoptosis, protected lymphoid and hematopoietic tissues (143), and reduced the incidence and severity of radiation-induced mucositis (108). A related compound, GC4419, is currently in Phase IIa testing (NCT01921426) in patients undergoing standard chemoradiation treatment of head and neck cancer, a setting in which up to 80% of patients develop severe oral mucositis.

# Mitochondria-targeted antioxidants

The direct and specific targeting of mitochondrial ROS formation by mitochondria-targeted antioxidants is another strategy still pursued to date. This will decrease mitochondrial ROS and prevent oxidative damage of important mitochondrial structures from ROS produced by extramitochondrial sources (*e.g.*, mitochondrial DNA is not well protected since efficient DNA repair systems do not exist in the matrix).

One of the first examples for this class of compounds was mitoquinone (1, 80), a quinone coupled to triphenylphosphonium, leading to significant accumulation in mitochondria. mitoTEMPO is another promising candidate for targeting mitochondrial ROS as it prevents adverse effects of angiotensin-II in experimental hypertension (30). A drawback of this strategy could be that these compounds require viable mitochondria with a certain membrane potential for uptake, which could limit their accumulation and accordingly antioxidant potency, especially in dysfunctional ROSproducing mitochondria. Several candidates of these mitochondria-targeted antioxidants are currently in latephase clinical trials (109, 138).

# Pharmacokinetic targeting

Another clinical approach to overcome the limitations or lack of evidence of chronic oral antioxidant therapy is the short-term infusion of antioxidants such as vitamin C. Positive reports include the reduction of histamine levels in allergy (45), enhanced hearing recovery in idiopathic sudden hearing loss (74), anti-inflammation improved recovery and lower rate of complication in pancreatitis (35), reduction of postzoster neuralgia (130), and reduction of gastrointestinal symptoms, fatigue, and pain in advanced cancer (22). The mechanistic rational is that such acute parenteral treatment targets massive inflammation-associated ROS generation and any side effects due to interference with physiological functions of ROS, such as in chronic oral therapy, are not observed.

A recent phase-I study investigating primarily safety of high-dose vitamin C infusion reported also positive effects on inflammation and endothelial function (39). However, endothelial function is only a surrogate parameter and hard outcome studies need to show true patient-relevant benefit. Prospective studies investigating the intravenous application of vitamin C in surgery and burn patients reported improved organ protection and function (83, 119, 142). On the other hand, preclinical studies prove that pharmacologic vitamin C concentrations are cytotoxic for cancer cell lines through the generation of  $H_2O_2$ .

There appears to be a multifactorial process where vitamin C induces cell cycle arrest, autophagy, apoptosis, and necrosis, depending on the cell genotype, the ascorbate concentration, and exposure time (113). The high (millimolar) concentrations necessary for this tumor cytotoxic effect can only be achieved intravenously because oral administration of vitamin C does not increase plasma levels above approx. 0.22 mM (84). There are currently three phase I/IIa studies being carried out in conjunction with the National Institutes of Health, which examine the effects of high-dosage vitamin C infusions adjuvant to standard therapy in the case of advanced tumors (93, 104, 152).

# **New Indirect Antioxidant Approaches**

A different concept to antioxidant therapy follows the proposition that it is not the antioxidant molecules, but the antioxidant enzymes that provide relevant therapeutic benefit. Molecules that augment the physiological antioxidant response without being antioxidants themselves are here termed indirect antioxidants and include NRF2 agonists and resveratrol.

### NRF2 activators

The transcription factor, nuclear factor (erythroid-derived 2)-like 2 (NRF2), is a master regulator of cell homeostasis that regulates the expression of antioxidant and cytoprotective genes that contain a specific enhancer sequence in their regulatory regions termed antioxidant response element (ARE). These genes account for more than 1% of the human genome and include antioxidant genes, *HMOX1* (coding heme oxygenase-1) and *NQO1* (coding NAD(P)H quinone oxidoreductase-1), and gene encoding enzymes involved in glutathione metabolism or in the generation of nicotinamide adenine dinucleotide phosphate (NADPH), *etc.* (Fig. 5A). For a detailed catalog of NRF2-regulated genes, see Hayes and Dinkova-Kostova (48).

Modulation of NRF2 activity may provide two advantages over direct antioxidants. First, the induction of NADPH, GSH, and thioredoxin metabolism is a natural system, resulting possibly in antioxidant activity in places where needed while leaving physiological ROS signaling intact. Second, because proteins have a longer half-life than small-molecule activators, the effect on the antioxidant defense may be more prolonged.

The main mechanism of regulation of NRF2 stability is its binding to the E3 ligase adapter kelch-like ECH-associated protein 1 (KEAP1) (Fig. 5B). This protein targets NRF2 for Cullin3/Rbx-mediated ubiquitination and subsequent proteasomal degradation. However, a drop in GSH concentration (characteristic of oxidant attack) or the presence of electrophiles leads to sulfhydryl bonding or adduct formation, respectively. This affects several cysteine residues in KEAP1, among which C151, C273, and C288 appear to be the most sensitive, probably because their pKa value is lower (pKa 4–5) compared with other cysteine residues (pKa 8.5) (21, 91). Such modification of KEAP1 leads to disruption of its interaction with NRF2, its stabilization, nuclear translocation, and activation of about 250 ARE-containing genes.

All NRF2 agonists modify the sulfhydryl group of cysteine residues of KEAP1 by oxidation or adduct formation. These molecules include allyl sulfides, dithiolethiones, flavonoids, isothiocyanates, polyphenols, and terpenoids (49). However, electrophilic compounds may also react with redox-sensitive cysteine residues present in the catalytic center of several phosphatases. This interaction leads to upregulation of signaling pathways that further impinge on NRF2 activation. For instance, carnosol (96), tert-butylhydroquinone (127), synthetic triterpenoids (2-cyano-3,12-dioxooleana-1,9-dien-28-oic acid-imidazolide [CDDO; CDDO-Im]) (120), or nordihydroguaiaretic acid (126) activate AKT signaling.

Several reports demonstrate that phosphatase and tensin homolog (PTEN), which is mutated in a large number of human tumors, is a redox-sensitive phosphatase (78). Importantly, the catalytic C124 of PTEN can be modified through adduct formation with strong electrophiles such as CDDO-Im (120) and tert-buthylhydroquinone (127). This modification results in loss of its lipid phosphatase activity and yields a more sustained activation of signaling events downstream of PI3K. These events include the activation of NRF2 by preventing its degradation through a KEAP1-independent mechanism that involves its phosphorylation by glycogen synthase-3 and further ubiquitination by a  $\beta$ -TrCP/Cullin1/Rbx complex (Fig. 5C) (122, 123).

In summary, when considering electrophilic targeting to NRF2, it is important to remember that not only KEAP1 but also other redox-sensitive enzymes may cooperate to regulate this antioxidant pathway.

The most compelling evidence in favor of a role of NRF2 in prevention or protection from disease has been gathered from results obtained with NRF2 knockout mice. With increasing age, these mice present with chronic pathologies related to oxidant and inflammatory stress, including cognitive deficits (66), depressive disorder (97), and increased incidence of lupus-like autoimmune nephritis (92). In humans, many single-nucleotide polymorphisms have been identified in the coding and noncoding regions of the gene encoding NRF2, *NFE2L2*, and epidemiological studies have revealed significant associations of *NFE2L2* haplotypes with risks in pulmonary, gastrointestinal, autoimmune, and neurodegenerative diseases (27).

The most successful case reported so far of an indirect antioxidant targeting NRF2 is the ester derivative of fumaric acid, dimethyl fumarate (DMF). DMF crosses the gastrointestinal barrier, after which it is converted into the active principle monomethyl fumarate (MMF), which binds KEAP1. This binding disrupts the KEAP1/NRF2 interaction and leads to upregulation of the transcriptional NRF2

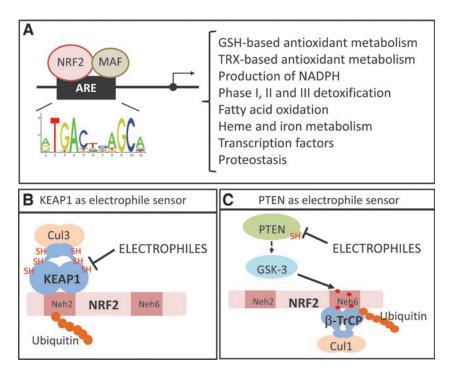


FIG. 5. Scheme on the antioxidant potential of the NRF2 signaling pathway and its pharmacologic targeting. (A) NRF2 is a basic region leucine zipper (bZip) transcription factor that makes dimers with proteins of the small MAF family. These heterodimers recognize an antioxidant response element (ARE) in about 250 genes (48). The consensus sequence is taken from JASPAR (99) (B) Regulation of NRF2/KEAP1 by electrophilic compounds. Under basal homeostatic conditions, NRF2 binds a KEAP1 homodimer through two tethering sites of low affinity (DLG) and high affinity (ETGE) located at the Neh2 domain. KEAP1 is a redox-sensitive E3 ligase adapter that leads to Cul3/Rbx-mediated ubiquitination of NRF2 and subsequent degradation by the proteasome. Adduct formation with electrophiles induces a conformational change in KEAP1 that prevents further degradation of NRF2 (145). (C) Regulation of NRF2/PTEN by electrophilic compounds. The lipid phosphatase activity of PTEN eliminates 3-phosphoinositides required to activate AKT, thus yielding glycogen synthase kinase-3 (GSK-3) active. GSK-3 phosphorylates NRF2 in several residues (DSGIS) of the Neh6 domain and creates a phosphodegron that is recognized by the E3 ligase adapter,  $\beta$ -TrCP. This process leads to a Cul1/Rbx-mediated ubiquitination and further proteasome degradation of NRF2. However, some electrophiles react with the critical C124 residue of PTEN located at the catalytic center to inhibit this enzyme. Then, 3-phosphoinositide levels remain high, leading to enhanced activation of AKT and inhibition of GSK-3. Under these conditions, NRF2 escapes GSK-3-mediated phosphorylation and recognition by the  $\beta$ -TrCP/Cul1/Rbx ubiquitin ligase (123, 127). AKT, protein kinase B; KEAP1, kelch-like ECH-associated protein 1; PTEN, phosphatase and tensin homolog. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

signature of antioxidant genes. Topical and oral administration of DMF (as a salt solution commercialized as Fumaderm) was indicated as early as year 1994 for the treatment of psoriasis before psoriasis was known to be an autoimmune disease or the molecular targets of DMF were identified (3).

The immunomodulatory effect of DMF has been exploited more recently for other autoimmune diseases such as lupus erythematosus (146), asthma, and arthritis (131), and a pharmaceutical formulation of DMF, termed FP187, is under development for use in autoimmune diseases.

DMF is being used in the treatment of relapsing—remitting multiple sclerosis (MS) (16). MMF is more potent than DMF in NRF2 activation, but the former is probably metabolized more rapidly. In an attempt to allow DMF to bypass gastric metabolism, it has been packaged in an oral delayed-release formulation known as BG-12. Following the success with BG-12 in two clinical trials, CONFIRM and DEFINE, this formulation has been commercialized as Tecfidera (16). Finally, a new formulation of MMF as a prodrug is currently in phase I trial for MS (61a).

Other lines of research have focused on targeting NRF2 in degenerative diseases with low-grade chronic inflammation using very potent synthetic triterpenoids that target KEAP1/NRF2. One of these compounds, CDDO-methyl ester (also known as bardoxolone methyl), was studied for therapy of diabetic nephropathy (151). The initial excitement about this compound was hampered by the observation that in the phase III clinical trial, termed BEACON, there was a small yet significant increase in heart failure in the treated arm compared with the placebo arm. This drawback was not related to NRF2 targeting, but rather to alteration of the endothelin signaling, leading to reduction in urine volume and sodium excretion in some patients with advanced chronic kidney disease (26). As a result, bardoxolone methyl is now being tested in pulmonary arterial hypertension (27a, 60), melanoma (54), and Friedreich's ataxia (27b). Moreover, its safety and efficacy for chronic kidney disease are being reevaluated (81a)

A third NRF2-activating principle is the isothiocyanate sulforaphane. Sulforaphane has been isolated from 3-day-old broccoli sprouts and other cruciferous vegetables as a product of the enzymatic cleavage of the glucosinolate, glucoraphanin, by the plant enzyme, myrosinase (37). Due to its electrophilic structure, sulforaphane interacts with specific

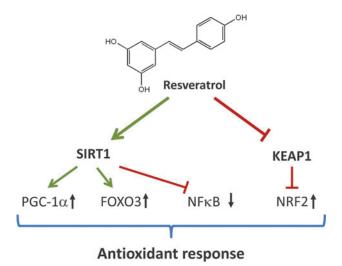


FIG. 6. Two mechanisms used by resveratrol to provide oxidant protection. SIRT1 is a histone deacetylase that participates in chromatin remodeling (not shown) and also in post-translational modification of transcription factors, such as PGC-1 $\alpha$ , FOXO3a, and NF- $\kappa$ B. In addition, it inhibits KEAP1 and leads to NRF2 activation. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

redox-sensitive cysteine residues in KEAP1, including Cys151. This disrupts the Keap1/NRF2 complex and leads to the increase in NRF2 protein levels and transcriptional activity (31). Most clinical studies performed with sulforaphane have used a highly standardized formulation of broccoli sprout extracts. In contrast to DMF and bardoxolone methyl, a drawback of sulforaphane is the absence of a pure formulation that could be used clinically and the lack of commercial value that could attract the strong investment that pipelines on drug development require. Nevertheless, several attempts are being made to provide proof of concept that NRF2 targeting with sulforaphane has a therapeutic value. Currently, at least 32 studies assess its clinical efficacy in chronic diseases such as cancer, asthma, autism, schizophrenia, chronic kidney disease, type 2 diabetes, and cystic fibrosis.

# Resveratrol

A comprehensive overview about resveratrol, including its targets and findings in clinical trials, is provided in a recent review (23) and schematized in Figure 6. The antioxidant phytochemical resveratrol was initially viewed as a direct ROS scavenger, but recent data suggest that its beneficial effects—if any—are rather mediated by indirect antioxidant mechanisms (85). Resveratrol increases the expression of antioxidant enzymes, such as SODs, catalase, glutathione peroxidase, and others, and reduces the expression of the ROS-forming NADPH oxidase type 4 (NOX4) (139).

In addition, resveratrol confers antioxidant activity by induction of the NRF2-heme oxygenase-1 pathway (69, 70) and it restores the activity of eNOS by increasing the synthesis of its cofactor, tetrahydrobiopterin (BH<sub>4</sub>) (155). There is also good evidence that resveratrol not only acts on gene expression *via* miRNAs as well as epigenetic modifications *via* activation of the NAD<sup>+</sup>-dependent deacetylase sirtuin 1

(SIRT1) (38, 85, 105) but also on proteins of the DNA repair machinery, thereby contributing to genome stability (41).

The most compelling clinical studies on the use of resveratrol in antioxidant therapy have demonstrated a reduction in redox biomarkers together with an increase in the NRF2 signature (42). In some cases, preliminary evidence of a salutary effect has been reported. Thus, resveratrol improves insulin sensitivity in patients with type 2 diabetes (19). In obese humans, resveratrol supplementation induced metabolic changes mimicking the effects of calorie restriction (144). Resveratrol demonstrated anti-inflammatory, antioxidant, and hypotriglyceridemic effects in healthy smokers (15). Further phase III studies will be required to determine real efficacy of this indirect antioxidant.

#### **Conclusions**

In view of the disparate results obtained with antioxidant compounds ranging from possibly beneficial to many futile to some harmful effects, it is necessary to reevaluate antioxidant therapy with a revised concept of oxidative stress that considers not only over ROS imbalance in a quantitative manner but also the molecular nature of ROS local cellular and tissue production and the enzymatic machinery in charge of its regulation.

While chronic, not indicated, therapy with antioxidants (especially lipid-soluble ones) must clearly not be recommended, there may be potential for the use of selective antioxidants, such as vitamin C, in situations of deficiencies or short-term overproduction accessible to parenteral high-dose therapy. Whether there is room for SOD mimetics needs to be shown.

More modern indirect antioxidants that target redox enzymes seem to be the more promising. NRF2 agonists may have the benefit of not causing reductive stress, but instead to upregulate endogenous antioxidant defense systems. Other approaches such as interfering with specific ROS-producing enzymes directly or even functionally repairing ROS-induced damage are discussed in another review of this Forum (see Targets review in the same Forum). Time and clinical trials investigating patient-relevant outcomes will tell.

# **Acknowledgments**

A.C. is the recipient of a research grant, SAF2013-43271-R, from the Spanish Ministry of Economy and Competitiveness. A.D. was supported by grants by the Federal Ministry of Education and Research to the Center of Thrombosis and Hemostasis Mainz (BMBF 01EO1003). R.S. is the recipient of a Senior Principal Research Fellowship of the National Health and Medical Research Council of Australia. Several authors of this review were supported by the European Cooperation in Science and Technology (COST Action BM1203/EU-ROS).

# **Author Disclosure Statement**

Claudia Vollbracht is a part-time employee of Pascoe Pharmazeutische Präparate GmbH, Giessen, Germany, which markets parenteral vitamin C products. Dennis Riley is the Chief Scientific Officer of Galera Therapeutics, Inc. (Malvern, PA), which is developing GC4419, an SOD mimetic. For all other authors, no competing financial interests exist.

#### References

1. Adlam VJ, Harrison JC, Porteous CM, James AM, Smith RA, Murphy MP, and Sammut IA. Targeting an antioxidant to mitochondria decreases cardiac ischemia-reperfusion injury. *FASEB J* 19: 1088–1095, 2005.

- Ali-Hassan-Sayegh S, Mirhosseini SJ, Rezaeisadrabadi M, Dehghan HR, Sedaghat-Hamedani F, Kayvanpour E, Popov AF, and Liakopoulos OJ. Antioxidant supplementations for prevention of atrial fibrillation after cardiac surgery: an updated comprehensive systematic review and meta-analysis of 23 randomized controlled trials. *Interact Cardiovasc Thorac Surg* 18: 646–654, 2014.
- Altmeyer PJ, Matthes U, Pawlak F, Hoffmann K, Frosch PJ, Ruppert P, Wassilew SW, Horn T, Kreysel HW, Lutz G, et al. Antipsoriatic effect of fumaric acid derivatives. Results of a multicenter double-blind study in 100 patients. J Am Acad Dermatol 30: 977–981, 1994.
- Arango D, Morohashi K, Yilmaz A, Kuramochi K, Parihar A, Brahimaj B, Grotewold E, and Doseff AI. Molecular basis for the action of a dietary flavonoid revealed by the comprehensive identification of apigenin human targets. *Proc Natl Acad Sci U S A* 110: E2153–E2162, 2013.
- Ashor AW, Lara J, Mathers JC, and Siervo M. Effect of vitamin C on endothelial function in health and disease: a systematic review and meta-analysis of randomised controlled trials. *Atherosclerosis* 235: 9–20, 2014.
- Aston K, Rath N, Naik A, Slomczynska U, Schall OF, and Riley DP. Computer-aided design (CAD) of Mn(II) complexes: superoxide dismutase mimetics with catalytic activity exceeding the native enzyme. *Inorg Chem* 40: 1779–1789, 2001.
- Batinic-Haberle I, Rajic Z, Tovmasyan A, Reboucas JS, Ye X, Leong KW, Dewhirst MW, Vujaskovic Z, Benov L, and Spasojevic I. Diverse functions of cationic Mn(III) Nsubstituted pyridylporphyrins, recognized as SOD mimics. Free Radic Biol Med 51: 1035–1053, 2011.
- 8. Batinic-Haberle I, Tovmasyan A, Roberts ER, Vujaskovic Z, Leong KW, and Spasojevic I. SOD therapeutics: latest insights into their structure-activity relationships and impact on the cellular redox-based signaling pathways. *Antioxid Redox Signal* 20: 2372–2415, 2014.
- Bauersachs J and Widder JD. Reductive stress: linking heat shock protein 27, glutathione, and cardiomyopathy? *Hypertension* 55: 1299–1300, 2010.
- Bell PG, McHugh MP, Stevenson E, and Howatson G. The role of cherries in exercise and health. *Scand J Med Sci Sports* 24: 477–490, 2014.
- Benfeito S, Oliveira C, Soares P, Fernandes C, Silva T, Teixeira J, and Borges F. Antioxidant therapy: still in search of the 'magic bullet'. *Mitochondrion* 13: 427–435, 2013.
- 12. Benhar M and Stamler JS. A central role for S-nitrosylation in apoptosis. *Nat Cell Biol* 7: 645–646, 2005.
- 13. Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, and Gluud C. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA* 297: 842–857, 2007.
- Bjelakovic G, Nikolova D, Simonetti RG, and Gluud C. Antioxidant supplements for prevention of gastrointestinal cancers: a systematic review and meta-analysis. *Lancet* 364: 1219–1228, 2004.
- Bo S, Ciccone G, Castiglione A, Gambino R, De Michieli F, Villois P, Durazzo M, Cavallo-Perin P, and Cassader

- M. Anti-inflammatory and antioxidant effects of resveratrol in healthy smokers a randomized, double-blind, placebo-controlled, cross-over trial. *Curr Med Chem* 20: 1323–1331, 2013.
- Bomprezzi R. Dimethyl fumarate in the treatment of relapsing-remitting multiple sclerosis: an overview. *Ther Adv Neurol Disord* 8: 20–30, 2015.
- Bosch J, Lonn E, Pogue J, Arnold JM, Dagenais GR, Yusuf S, and Investigators HH-TS. Long-term effects of ramipril on cardiovascular events and on diabetes: results of the HOPE study extension. *Circulation* 112: 1339– 1346, 2005.
- Braakhuis AJ and Hopkins WG. Impact of dietary antioxidants on sport performance: a review. Sports Med 45: 939–955, 2015.
- 19. Brasnyo P, Molnar GA, Mohas M, Marko L, Laczy B, Cseh J, Mikolas E, Szijarto IA, Merei A, Halmai R, Meszaros LG, Sumegi B, and Wittmann I. Resveratrol improves insulin sensitivity, reduces oxidative stress and activates the Akt pathway in type 2 diabetic patients. *Br J Nutr* 106: 383–389, 2011.
- Brown DI and Griendling KK. Regulation of signal transduction by reactive oxygen species in the cardiovascular system. Circ Res 116: 531–549, 2015.
- Bryan HK, Olayanju A, Goldring CE, and Park BK. The Nrf2 cell defence pathway: Keap1-dependent and -independent mechanisms of regulation. *Biochem Pharmacol* 85: 705–717, 2013.
- Carr AC, Vissers MC, and Cook JS. The effect of intravenous vitamin C on cancer- and chemotherapy-related fatigue and quality of life. Front Oncol 4: 283, 2014.
- Carrizzo A, Forte M, Damato A, Trimarco V, Salzano F, Bartolo M, Maciag A, Puca AA, and Vecchione C. Antioxidant effects of resveratrol in cardiovascular, cerebral and metabolic diseases. *Food Chem Toxicol* 61: 215–226, 2013.
- Chen AF, Chen DD, Daiber A, Faraci FM, Li H, Rembold CM, and Laher I. Free radical biology of the cardiovascular system. *Clin Sci (Lond)* 123: 73–91, 2012.
- Chen GC, Lu DB, Pang Z, and Liu QF. Vitamin C intake, circulating vitamin C and risk of stroke: a meta-analysis of prospective studies. J Am Heart Assoc 2: e000329, 2013.
- 26. Chin MP, Reisman SA, Bakris GL, O'Grady M, Linde PG, McCullough PA, Packham D, Vaziri ND, Ward KW, Warnock DG, and Meyer CJ. Mechanisms contributing to adverse cardiovascular events in patients with type 2 diabetes mellitus and stage 4 chronic kidney disease treated with bardoxolone methyl. Am J Nephrol 39: 499–508, 2014.
- Cho HY. Genomic structure and variation of nuclear factor (erythroid-derived 2)-like 2. Oxid Med Cell Longev 2013: 286524, 2013.
- 27a. Clinicaltrials.gov. 2015. Bardoxolone Methyl Evaluation in Patients With Pulmonary Arterial Hypertension (PAH)— LARIAT. https://clinicaltrials.gov/ct2/show/NCT02036970. Accessed August 2015.
- 27b. Clinicaltrials.gov. 2015. RTA 408 Capsules in Patients With Friedreich's Ataxia–MOXIe. https://clinicaltrials.gov/ct2/show/NCT02255435. Accessed August 2015.
- 28. Cynshi O, Kawabe Y, Suzuki T, Takashima Y, Kaise H, Nakamura M, Ohba Y, Kato Y, Tamura K, Hayasaka A, Higashida A, Sakaguchi H, Takeya M, Takahashi K, Inoue K, Noguchi N, Niki E, and Kodama T. Antiatherogenic effects of the antioxidant BO-653 in three different animal models. *Proc Natl Acad Sci U S A* 95: 10123–10128, 1998.

- 29. Daiber A, Zou MH, Bachschmid M, and Ullrich V. Ebselen as a peroxynitrite scavenger *in vitro* and *ex vivo*. *Biochem Pharmacol* 59: 153–160, 2000.
- Dikalova AE, Bikineyeva AT, Budzyn K, Nazarewicz RR, McCann L, Lewis W, Harrison DG, and Dikalov SI. Therapeutic targeting of mitochondrial superoxide in hypertension. *Circ Res* 107: 106–116, 2010.
- Dinkova-Kostova AT and Talalay P. Direct and indirect antioxidant properties of inducers of cytoprotective proteins. Mol Nutr Food Res 52 Suppl 1: S128–S138, 2008.
- 32. Donko A, Morand S, Korzeniowska A, Boudreau HE, Zana M, Hunyady L, Geiszt M, and Leto TL. Hypothyroidism-associated missense mutation impairs NADPH oxidase activity and intracellular trafficking of Duox2. Free Radic Biol Med 73: 190–200, 2014.
- 33. Dotan Y, Pinchuk I, Lichtenberg D, and Leshno M. Decision analysis supports the paradigm that indiscriminate supplementation of vitamin E does more harm than good. *Arterioscler Thromb Vasc Biol* 29: 1304–1309, 2009.
- 34. Drummond GR, Cai H, Davis ME, Ramasamy S, and Harrison DG. Transcriptional and posttranscriptional regulation of endothelial nitric oxide synthase expression by hydrogen peroxide. *Circ Res* 86: 347–354, 2000.
- Du WD, Yuan ZR, Sun J, Tang JX, Cheng AQ, Shen DM, Huang CJ, Song XH, Yu XF, and Zheng SB. Therapeutic efficacy of high-dose vitamin C on acute pancreatitis and its potential mechanisms. World J Gastroenterol 9: 2565–2569, 2003.
- Eidelman RS, Hollar D, Hebert PR, Lamas GA, and Hennekens CH. Randomized trials of vitamin E in the treatment and prevention of cardiovascular disease. *Arch Intern Med* 164: 1552–1556, 2004.
- Fahey JW, Zhang Y, and Talalay P. Broccoli sprouts: an exceptionally rich source of inducers of enzymes that protect against chemical carcinogens. *Proc Natl Acad Sci* U S A 94: 10367–10372, 1997.
- Farghali H, Kutinova Canova N, and Lekic N. Resveratrol and related compounds as antioxidants with an allosteric mechanism of action in epigenetic drug targets. *Physiol Res* 62: 1–13, 2013.
- 39. Fowler AA, 3rd, Syed AA, Knowlson S, Sculthorpe R, Farthing D, Dewilde C, Farthing CA, Larus TL, Martin E, Brophy DF, Gupta S, Fisher BJ, and Natarajan R. Phase I safety trial of intravenous ascorbic acid in patients with severe sepsis. *J Transl Med* 12: 32, 2014.
- 40. Fox JT, Sakamuru S, Huang R, Teneva N, Simmons SO, Xia M, Tice RR, Austin CP, and Myung K. High-throughput genotoxicity assay identifies antioxidants as inducers of DNA damage response and cell death. *Proc Natl Acad Sci U S A* 109: 5423–5428, 2012.
- 41. Gatz SA and Wiesmuller L. Take a break—resveratrol in action on DNA. *Carcinogenesis* 29: 321–332, 2008.
- 42. Ghanim H, Sia CL, Abuaysheh S, Korzeniewski K, Patnaik P, Marumganti A, Chaudhuri A, and Dandona P. An antiinflammatory and reactive oxygen species suppressive effects of an extract of Polygonum cuspidatum containing resveratrol. *J Clin Endocrinol Metab* 95: E1– E8 2010
- 43. Griendling KK and FitzGerald GA. Oxidative stress and cardiovascular injury: Part II: animal and human studies. *Circulation* 108: 2034–2040, 2003.
- 44. Gustafson CB, Yang C, Dickson KM, Shao H, Van Booven D, Harbour JW, Liu ZJ, and Wang G. Epigenetic reprogramming of melanoma cells by vitamin C treatment. *Clin Epigenetics* 7: 51, 2015.

- 45. Hagel AF, Layritz CM, Hagel WH, Hagel HJ, Hagel E, Dauth W, Kressel J, Regnet T, Rosenberg A, Neurath MF, Molderings GJ, and Raithel M. Intravenous infusion of ascorbic acid decreases serum histamine concentrations in patients with allergic and non-allergic diseases. *Naunyn Schmiedebergs Arch Pharmacol* 386: 789–793, 2013.
- 46. Halliwell B. Cell culture, oxidative stress, and antioxidants: avoiding pitfalls. *Biomed J* 37: 99–105, 2014.
- 47. Harris HR, Orsini N, and Wolk A. Vitamin C and survival among women with breast cancer: a meta-analysis. *Eur J Cancer* 50: 1223–1231, 2014.
- Hayes JD and Dinkova-Kostova AT. The Nrf2 regulatory network provides an interface between redox and intermediary metabolism. *Trends Biochem Sci* 39: 199–218, 2014.
- Hayes JD, McMahon M, Chowdhry S, and Dinkova-Kostova AT. Cancer chemoprevention mechanisms mediated through the Keap1-Nrf2 pathway. *Antioxid Redox* Signal 13: 1713–1748, 2010.
- 50. Heitzer T, Finckh B, Albers S, Krohn K, Kohlschutter A, and Meinertz T. Beneficial effects of alpha-lipoic acid and ascorbic acid on endothelium-dependent, nitric oxide-mediated vasodilation in diabetic patients: relation to parameters of oxidative stress. *Free Radic Biol Med* 31: 53–61, 2001.
- Hemila H. Vitamin C may alleviate exercise-induced bronchoconstriction: a meta-analysis. *BMJ Open* 3: e002416, 2013.
- Hemila H and Chalker E. Vitamin C for preventing and treating the common cold. *Cochrane Database Syst Rev* 1: CD000980, 2013.
- Henry PT and Chandy MJ. Effect of ascorbic acid on infarct size in experimental focal cerebral ischaemia and reperfusion in a primate model. *Acta Neurochir (Wien)* 140: 977–980, 1998.
- 54. Hong DS, Kurzrock R, Supko JG, He X, Naing A, Wheler J, Lawrence D, Eder JP, Meyer CJ, Ferguson DA, Mier J, Konopleva M, Konoplev S, Andreeff M, Kufe D, Lazarus H, Shapiro GI, and Dezube BJ. A phase I first-in-human trial of bardoxolone methyl in patients with advanced solid tumors and lymphomas. Clin Cancer Res 18: 3396–3406, 2012.
- 55. This reference has been deleted.
- 56. This reference has been deleted.
- 57. This reference has been deleted.
- 58. This reference has been deleted.
- 59. This reference has been deleted.
- Hu J, Xu Q, McTiernan C, Lai YC, Osei-Hwedieh D, and Gladwin M. Novel targets of drug treatment for pulmonary hypertension. *Am J Cardiovasc Drugs* 15: 225–234, 2015.
- 61. Hunt JA, Lee J, and Groves JT. Amphiphilic peroxynitrite decomposition catalysts in liposomal assemblies. *Chem Biol* 4: 845–858, 1997.
- 61a. Hunt TL, Durairaj C, Leigh-Pemberton R, Jiang Y, Manthis J, and Hard M. 2015 Safety, Tolerability, and Pharmacokinetics of ALKS 8700, a Novel Oral Therapy for Relapsing-RemittingMultiple Sclerosis, in Healthy Subjects. The 2015 Annual Meeting of the Consortium of Multiple Sclerosis Centers (CMSC). http://annualmeeting .mscare.org/ Accessed August 2015.
- 61b. Huntington Study Group. 2014. Announcement of 2CARE Early Study Closure. http://huntingtonstudygroup.org/tag/2care/. Accessed August 2015.
- 62. Ischiropoulos H and Beckman JS. Oxidative stress and nitration in neurodegeneration: cause, effect, or association? *J Clin Invest* 111: 163–169, 2003.

63. Ishihara O, Hayashi M, Osawa H, Kobayashi K, Takeda S, Vessby B, and Basu S. Isoprostanes, prostaglandins and tocopherols in pre-eclampsia, normal pregnancy and nonpregnancy. *Free Radic Res* 38: 913–918, 2004.

- 64. Jackson MJ. Redox regulation of adaptive responses in skeletal muscle to contractile activity. *Free Radic Biol Med* 47: 1267–1275, 2009.
- Ji LL. Antioxidant signaling in skeletal muscle: a brief review. Exp Gerontol 42: 582–593, 2007.
- 66. Jo C, Gundemir S, Pritchard S, Jin YN, Rahman I, and Johnson GV. Nrf2 reduces levels of phosphorylated tau protein by inducing autophagy adaptor protein NDP52. *Nat Commun* 5: 3496, 2014.
- 67. Johnson BD, Mather KJ, Newcomer SC, Mickleborough TD, and Wallace JP. Vitamin C prevents the acute decline of flow-mediated dilation after altered shear rate patterns. *Appl Physiol Nutr Metab* 38: 268–274, 2013.
- Jones DP and Go YM. Redox compartmentalization and cellular stress. *Diabetes Obes Metab* 12 Suppl 2: 116–125, 2010.
- 69. Juan SH, Cheng TH, Lin HC, Chu YL, and Lee WS. Mechanism of concentration-dependent induction of heme oxygenase-1 by resveratrol in human aortic smooth muscle cells. *Biochem Pharmacol* 69: 41–48, 2005.
- Kaga S, Zhan L, Matsumoto M, and Maulik N. Resveratrol enhances neovascularization in the infarcted rat myocardium through the induction of thioredoxin-1, heme oxygenase-1 and vascular endothelial growth factor. *J Mol Cell Cardiol* 39: 813–822, 2005.
- 71. Kaiser J. Biomedical research. Antioxidants could spur tumors by acting on cancer gene. *Science* 343: 477, 2014.
- 72. Kajimoto H, Hashimoto K, Bonnet SN, Haromy A, Harry G, Moudgil R, Nakanishi T, Rebeyka I, Thebaud B, Michelakis ED, and Archer SL. Oxygen activates the Rho/Rho-kinase pathway and induces RhoB and ROCK-1 expression in human and rabbit ductus arteriosus by increasing mitochondria-derived reactive oxygen species: a newly recognized mechanism for sustaining ductal constriction. Circulation 115: 1777–1788, 2007.
- Kamat CD, Gadal S, Mhatre M, Williamson KS, Pye QN, and Hensley K. Antioxidants in central nervous system diseases: preclinical promise and translational challenges. *J Alzheimers Dis* 15: 473–493, 2008.
- 74. Kang HS, Park JJ, Ahn SK, Hur DG, and Kim HY. Effect of high dose intravenous vitamin C on idiopathic sudden sensorineural hearing loss: a prospective single-blind randomized controlled trial. *Eur Arch Otorhinolaryngol* 270: 2631–2636, 2013.
- Karbach S, Wenzel P, Waisman A, Munzel T, and Daiber A. eNOS uncoupling in cardiovascular diseases—the role of oxidative stress and inflammation. *Curr Pharm Des* 20: 3579–3594, 2014.
- Kemp M, Go YM, and Jones DP. Nonequilibrium thermodynamics of thiol/disulfide redox systems: a perspective on redox systems biology. *Free Radic Biol Med* 44: 921–937, 2008.
- 77. Khaw KT, Bingham S, Welch A, Luben R, Wareham N, Oakes S, and Day N. Relation between plasma ascorbic acid and mortality in men and women in EPIC-Norfolk prospective study: a prospective population study. European Prospective Investigation into Cancer and Nutrition. *Lancet* 357: 657–663, 2001.
- Kitagishi Y and Matsuda S. Redox regulation of tumor suppressor PTEN in cancer and aging (Review). *Int J Mol Med* 31: 511–515, 2013.

 Kojo S. Oxygen is the key factor associated with the difference between *in vivo* and *in vitro* effects of antioxidants. *Proc Natl Acad Sci U S A* 109: E2028; author reply E2029, 2012.

- 80. Koopman WJ, Verkaart S, Visch HJ, van der Westhuizen FH, Murphy MP, van den Heuvel LW, Smeitink JA, and Willems PH. Inhibition of complex I of the electron transport chain causes O2-. -mediated mitochondrial outgrowth. *Am J Physiol Cell Physiol* 288: C1440–C1450, 2005.
- 81. Kris-Etherton PM, Lichtenstein AH, Howard BV, Steinberg D, Witztum JL; Nutrition Committee of the American Heart Association Council on Nutrition, Physical Activity, and Metabolism. Antioxidant vitamin supplements and cardiovascular disease. *Circulation* 110: 637–641, 2004.
- 81a. Kyowa Hakko Kirin Co, ltd. 2013. Kyowa Hakko Kirin Announces the Development Status of Bardoxolone Methyl (RTA 402) in Patients with Chronic Kidney Disease and Type 2 Diabetes in Japan. www.kyowa-kirin.com/ news\_releases/2013/e20131111\_01.html (Accessed August 2015).
- 82. Lee DH, Folsom AR, Harnack L, Halliwell B, and Jacobs DR, Jr. Does supplemental vitamin C increase cardio-vascular disease risk in women with diabetes? *Am J Clin Nutr* 80: 1194–1200, 2004.
- 83. Lee JY, Kim CJ, and Chung MY. Effect of high-dose vitamin C on oxygen free radical production and myocardial enzyme after tourniquet ischaemia-reperfusion injury during bilateral total knee replacement. *J Int Med Res* 38: 1519–1529, 2010.
- 84. Levine M, Padayatty SJ, and Espey MG. Vitamin C: a concentration-function approach yields pharmacology and therapeutic discoveries. *Adv Nutr* 2: 78–88, 2011.
- Li H, Xia N, and Forstermann U. Cardiovascular effects and molecular targets of resveratrol. *Nitric Oxide* 26: 102– 110, 2012.
- Linster CL and Van Schaftingen E. Vitamin C. Biosynthesis, recycling and degradation in mammals. FEBS J 274: 1–22, 2007.
- Liu XH, Xu CY, and Fan GH. Efficacy of N-acetylcysteine in preventing atrial fibrillation after cardiac surgery: a metaanalysis of published randomized controlled trials. *BMC Cardiovasc Disord* 14: 52, 2014.
- 88. Lobysheva I, Rath G, Sekkali B, Bouzin C, Feron O, Gallez B, Dessy C, and Balligand JL. Moderate caveolin-1 down-regulation prevents NADPH oxidase-dependent endothelial nitric oxide synthase uncoupling by angiotensin II in endothelial cells. *Arterioscler Thromb Vasc Biol* 31: 2098–2105, 2011.
- 89. Lonn E, Bosch J, Yusuf S, Sheridan P, Pogue J, Arnold JM, Ross C, Arnold A, Sleight P, Probstfield J, and Dagenais GR. Effects of long-term vitamin E supplementation on cardiovascular events and cancer: a randomized controlled trial. *JAMA* 293: 1338–1347, 2005.
- Lonn ME, Dennis JM, and Stocker R. Actions of "anti-oxidants" in the protection against atherosclerosis. Free Radic Biol Med 53: 863–884, 2012.
- 91. Ma Q. Role of nrf2 in oxidative stress and toxicity. *Annu Rev Pharmacol Toxicol* 53: 401–426, 2013.
- Ma Q, Battelli L, and Hubbs AF. Multiorgan autoimmune inflammation, enhanced lymphoproliferation, and impaired homeostasis of reactive oxygen species in mice lacking the antioxidant-activated transcription factor Nrf2. Am J Pathol 168: 1960–1974, 2006.
- Ma Y, Chapman J, Levine M, Polireddy K, Drisko J, and Chen Q. High-dose parenteral ascorbate enhanced

- chemosensitivity of ovarian cancer and reduced toxicity of chemotherapy. *Sci Transl Med* 6: 222ra18, 2014.
- 94. Makanae Y, Kawada S, Sasaki K, Nakazato K, and Ishii N. Vitamin C administration attenuates overload-induced skeletal muscle hypertrophy in rats. *Acta Physiol (Oxf)* 208: 57–65, 2013.
- 95. Mann JF, Lonn EM, Yi Q, Gerstein HC, Hoogwerf BJ, Pogue J, Bosch J, Dagenais GR, and Yusuf S. Effects of vitamin E on cardiovascular outcomes in people with mild-to-moderate renal insufficiency: results of the HOPE study. *Kidney Int* 65: 1375–1380, 2004.
- 96. Martin D, Rojo AI, Salinas M, Diaz R, Gallardo G, Alam J, De Galarreta CM, and Cuadrado A. Regulation of heme oxygenase-1 expression through the phosphatidylinositol 3-kinase/Akt pathway and the Nrf2 transcription factor in response to the antioxidant phytochemical carnosol. *J Biol Chem* 279: 8919–8929, 2004.
- 97. Martin-de-Saavedra MD, Budni J, Cunha MP, Gomez-Rangel V, Lorrio S, Del Barrio L, Lastres-Becker I, Parada E, Tordera RM, Rodrigues AL, Cuadrado A, and Lopez MG. Nrf2 participates in depressive disorders through an anti-inflammatory mechanism. *Psychoneuroendocrinology* 38: 2010–2022, 2013.
- 98. Masumoto H and Sies H. The reaction of ebselen with peroxynitrite. *Chem Res Toxicol* 9: 262–267, 1996.
- 99. Mathelier A, Zhao X, Zhang AW, Parcy F, Worsley-Hunt R, Arenillas DJ, Buchman S, Chen CY, Chou A, Ienasescu H, Lim J, Shyr C, Tan G, Zhou M, Lenhard B, Sandelin A, and Wasserman WW. JASPAR 2014: an extensively expanded and updated open-access database of transcription factor binding profiles. *Nucleic Acids Res* 42: D142–D147, 2014.
- 100. Matoba T, Shimokawa H, Nakashima M, Hirakawa Y, Mukai Y, Hirano K, Kanaide H, and Takeshita A. Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in mice. J Clin Invest 106: 1521–1530, 2000.
- 101. Medved I, Brown MJ, Bjorksten AR, Murphy KT, Petersen AC, Sostaric S, Gong X, and McKenna MJ. Nacetylcysteine enhances muscle cysteine and glutathione availability and attenuates fatigue during prolonged exercise in endurance-trained individuals. *J Appl Physiol* (1985) 97: 1477–1485, 2004.
- 102. Miller ER, 3rd, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, and Guallar E. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Ann Intern Med* 142: 37–46, 2005.
- 103. Mofarrahi M, Brandes RP, Gorlach A, Hanze J, Terada LS, Quinn MT, Mayaki D, Petrof B, and Hussain SN. Regulation of proliferation of skeletal muscle precursor cells by NADPH oxidase. *Antioxid Redox Signal* 10: 559– 574, 2008.
- 104. Monti DA, Mitchell E, Bazzan AJ, Littman S, Zabrecky G, Yeo CJ, Pillai MV, Newberg AB, Deshmukh S, and Levine M. Phase I evaluation of intravenous ascorbic acid in combination with gemcitabine and erlotinib in patients with metastatic pancreatic cancer. PLoS One 7: e29794, 2012.
- 105. Mukhopadhyay P, Pacher P, and Das DK. MicroRNA signatures of resveratrol in the ischemic heart. Ann N Y Acad Sci 1215: 109–116, 2011.
- 106. Muller A, Cadenas E, Graf P, and Sies H. A novel biologically active seleno-organic compound—I. Glutathione peroxidase-like activity in vitro and antioxidant capacity of PZ 51 (Ebselen). Biochem Pharmacol 33: 3235–3239, 1984.
- Muntwyler J, Hennekens CH, Manson JE, Buring JE, and Gaziano JM. Vitamin supplement use in a low-risk popu-

- lation of US male physicians and subsequent cardiovascular mortality. *Arch Intern Med* 162: 1472–1476, 2002.
- 108. Murphy CK, Fey EG, Watkins BA, Wong V, Rothstein D, and Sonis ST. Efficacy of superoxide dismutase mimetic M40403 in attenuating radiation-induced oral mucositis in hamsters. Clin Cancer Res 14: 4292–4297, 2008.
- 109. Murphy MP. Antioxidants as therapies: can we improve on nature? *Free Radic Biol Med* 66: 20–23, 2014.
- 110. Nezis IP and Stenmark H. p62 at the interface of autophagy, oxidative stress signaling, and cancer. *Antioxid Redox Signal* 17: 786–793, 2012.
- 111. Paffenholz R, Bergstrom RA, Pasutto F, Wabnitz P, Munroe RJ, Jagla W, Heinzmann U, Marquardt A, Bareiss A, Laufs J, Russ A, Stumm G, Schimenti JC, and Bergstrom DE. Vestibular defects in head-tilt mice result from mutations in Nox3, encoding an NADPH oxidase. *Genes Dev* 18: 486–491, 2004.
- 112. Palm M, Axelsson O, Wernroth L, and Basu S. F(2)-isoprostanes, tocopherols and normal pregnancy. *Free Radic Res* 43: 546–552, 2009.
- 113. Parrow NL, Leshin JA, and Levine M. Parenteral ascorbate as a cancer therapeutic: a reassessment based on pharmacokinetics. *Antioxid Redox Signal* 19: 2141–2156, 2013.
- 114. Paschalis V, Theodorou AA, Kyparos A, Dipla K, Zafeiridis A, Panayiotou G, Vrabas IS, and Nikolaidis MG. Low vitamin C values are linked with decreased physical performance and increased oxidative stress: reversal by vitamin C supplementation. *Eur J Nutr* 2014 [Epub ahead of print]; DOI: 10.1007/s00394-014-0821-x.
- 115. Pasternack RF and Skowronek WR, Jr. Catalysis of the disproportionation of superoxide by metalloporphyrins. J Inorg Biochem 11: 261–267, 1979.
- 116. Paulsen G, Cumming KT, Holden G, Hallen J, Ronnestad BR, Sveen O, Skaug A, Paur I, Bastani NE, Ostgaard HN, Buer C, Midttun M, Freuchen F, Wiig H, Ulseth ET, Garthe I, Blomhoff R, Benestad HB, and Raastad T. Vitamin C and E supplementation hampers cellular adaptation to endurance training in humans: a double-blind, randomised, controlled trial. *J Physiol* 592: 1887–1901, 2014.
- 117. Paulsen G, Hamarsland H, Cumming KT, Johansen RE, Hulmi JJ, Borsheim E, Wiig H, Garthe I, and Raastad T. Vitamin C and E supplementation alters protein signalling after a strength training session, but not muscle growth during 10 weeks of training. *J Physiol* 592: 5391–5408, 2014.
- 118. Perera RM and Bardeesy N. Cancer: when antioxidants are bad. *Nature* 475: 43–44, 2011.
- 119. Pignatelli P, Tanzilli G, Carnevale R, Di Santo S, Loffredo L, Celestini A, Proietti M, Tovaglia P, Mangieri E, Basili S, and Violi F. Ascorbic acid infusion blunts CD40L upregulation in patients undergoing coronary stent. *Cardiovasc Ther* 29: 385–394, 2011.
- 120. Pitha-Rowe I, Liby K, Royce D, and Sporn M. Synthetic triterpenoids attenuate cytotoxic retinal injury: cross-talk between Nrf2 and PI3K/AKT signaling through inhibition of the lipid phosphatase PTEN. *Invest Ophthalmol Vis Sci* 50: 5339–5347, 2009.
- 121. Poston L, Briley AL, Seed PT, Kelly FJ, Shennan AH; Vitamins in Pre-eclampsia (VIP) Trial Consortium. Vitamin C and vitamin E in pregnant women at risk for pre-eclampsia (VIP trial): randomised placebo-controlled trial. *Lancet* 367: 1145–1154, 2006.
- 122. Rada P, Rojo AI, Chowdhry S, McMahon M, Hayes JD, and Cuadrado A. SCF/{beta}-TrCP promotes glycogen synthase kinase 3-dependent degradation of the Nrf2

transcription factor in a Keap1-independent manner. *Mol Cell Biol* 31: 1121–1133, 2011.

- 123. Rada P, Rojo AI, Evrard-Todeschi N, Innamorato NG, Cotte A, Jaworski T, Tobon-Velasco JC, Devijver H, Garcia-Mayoral MF, Van Leuven F, Hayes JD, Bertho G, and Cuadrado A. Structural and functional characterization of Nrf2 degradation by the glycogen synthase kinase 3/beta-TrCP axis. *Mol Cell Biol* 32: 3486–3499, 2012.
- 124. Ray PD, Huang BW, and Tsuji Y. Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal* 24: 981–990, 2012.
- 125. Ristow M, Zarse K, Oberbach A, Kloting N, Birringer M, Kiehntopf M, Stumvoll M, Kahn CR, and Bluher M. Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc Natl Acad Sci U S A* 106: 8665–8670, 2009.
- 126. Rojo AI, Medina-Campos ON, Rada P, Zuniga-Toala A, Lopez-Gazcon A, Espada S, Pedraza-Chaverri J, and Cuadrado A. Signaling pathways activated by the phytochemical nordihydroguaiaretic acid contribute to a Keap1independent regulation of Nrf2 stability: role of glycogen synthase kinase-3. Free Radic Biol Med 52: 473–487, 2012.
- 127. Rojo AI, Rada P, Mendiola M, Ortega-Molina A, Wojdyla K, Rogowska-Wrzesinska A, Hardisson D, Serrano M, and Cuadrado A. The PTEN/NRF2 axis promotes human carcinogenesis. *Antioxid Redox Signal* 21: 2498–2514, 2014.
- Salvemini D and Riley DP. Nonpeptidyl mimetics of superoxide dismutase in clinical therapies for diseases. *Cell Mol Life Sci* 57: 1489–1492, 2000.
- Sayin VI, Ibrahim MX, Larsson E, Nilsson JA, Lindahl P, and Bergo MO. Antioxidants accelerate lung cancer progression in mice. Sci Transl Med 6: 221ra15, 2014.
- 130. Schencking M, Vollbracht C, Weiss G, Lebert J, Biller A, Goyvaerts B, and Kraft K. Intravenous Vitamin C in the treatment of shingles: results of a multicenter prospective cohort study. *Med Sci Monit* 18: CR215–CR224, 2012.
- Seidel P and Roth M. Anti-inflammatory dimethylfumarate: a potential new therapy for asthma? *Mediators Inflamm* 2013: 875403, 2013.
- 132. Sesso HD, Buring JE, Christen WG, Kurth T, Belanger C, MacFadyen J, Bubes V, Manson JE, Glynn RJ, and Gaziano JM. Vitamins E and C in the prevention of cardio-vascular disease in men: the Physicians' Health Study II randomized controlled trial. *JAMA* 300: 2123–2133, 2008.
- 133. Shekelle PG, Morton SC, Jungvig LK, Udani J, Spar M, Tu W, M JS, Coulter I, Newberry SJ, and Hardy M. Effect of supplemental vitamin E for the prevention and treatment of cardiovascular disease. *J Gen Intern Med* 19: 380–389, 2004.
- 134. Shen Y, Cai W, Lei S, and Zhang Z. Effect of high/low dose N-acetylcysteine on chronic obstructive pulmonary disease: a systematic review and meta-analysis. COPD 11: 351–358, 2014.
- 135. Shuaib A, Lees KR, Lyden P, Grotta J, Davalos A, Davis SM, Diener HC, Ashwood T, Wasiewski WW, Emeribe U; SAINT II Trial Investigators. NXY-059 for the treatment of acute ischemic stroke. N Engl J Med 357: 562–571, 2007.
- 136. Sies H. Oxidative stress: a concept in redox biology and medicine. *Redox Biol* 4: 180–183, 2015.
- Sleight P. The HOPE Study (Heart Outcomes Prevention Evaluation). J Renin Angiotensin Aldosterone Syst 1: 18– 20, 2000.
- Smith RA, Hartley RC, and Murphy MP. Mitochondriatargeted small molecule therapeutics and probes. *Antioxid Redox Signal* 15: 3021–3038, 2011.

139. Spanier G, Xu H, Xia N, Tobias S, Deng S, Wojnowski L, Forstermann U, and Li H. Resveratrol reduces endothelial oxidative stress by modulating the gene expression of superoxide dismutase 1 (SOD1), glutathione peroxidase 1 (GPx1) and NADPH oxidase subunit (Nox4). *J Physiol Pharmacol* 60 Suppl 4: 111–116, 2009.

- 140. Stacey MM, Vissers MC, and Winterbourn CC. Oxidation of 2-cys peroxiredoxins in human endothelial cells by hydrogen peroxide, hypochlorous acid, and chloramines. *Antioxid Redox Signal* 17: 411–421, 2012.
- 141. Stocker R. The ambivalence of vitamin E in atherogenesis. *Trends Biochem Sci* 24: 219–223, 1999.
- 142. Tanaka H, Matsuda T, Miyagantani Y, Yukioka T, Matsuda H, and Shimazaki S. Reduction of resuscitation fluid volumes in severely burned patients using ascorbic acid administration: a randomized, prospective study. *Arch Surg* 135: 326–331, 2000.
- 143. Thompson JS, Chu Y, Glass J, Tapp AA, and Brown SA. The manganese superoxide dismutase mimetic, M40403, protects adult mice from lethal total body irradiation. *Free Radic Res* 44: 529–540, 2010.
- 144. Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, Hoeks J, van der Krieken S, Ryu D, Kersten S, Moonen-Kornips E, Hesselink MK, Kunz I, Schrauwen-Hinderling VB, Blaak EE, Auwerx J, and Schrauwen P. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metab* 14: 612– 622, 2011.
- 145. Tong KI, Katoh Y, Kusunoki H, Itoh K, Tanaka T, and Yamamoto M. Keap1 recruits Neh2 through binding to ETGE and DLG motifs: characterization of the two-site molecular recognition model. *Mol Cell Biol* 26: 2887– 2900, 2006.
- 146. Tsianakas A, Herzog S, Landmann A, Patsinakidis N, Perusquia Ortiz AM, Bonsmann G, Luger TA, and Kuhn A. Successful treatment of discoid lupus erythematosus with fumaric acid esters. J Am Acad Dermatol 71: e15–e17, 2014.
- 147. Vanden Berghe T, Linkermann A, Jouan-Lanhouet S, Walczak H, and Vandenabeele P. Regulated necrosis: the expanding network of non-apoptotic cell death pathways. *Nat Rev Mol Cell Biol* 15: 135–147, 2014.
- 148. Venditti P, Napolitano G, Barone D, and Di Meo S. Effect of training and vitamin E administration on rat liver oxidative metabolism. *Free Radic Res* 48: 322–332, 2014.
- 149. Venturelli S, Sinnberg TW, Berger A, Noor S, Levesque MP, Bocker A, Niessner H, Lauer UM, Bitzer M, Garbe C, and Busch C. Epigenetic impacts of ascorbate on human metastatic melanoma cells. Front Oncol 4: 227, 2014.
- 150. Vivekananthan DP, Penn MS, Sapp SK, Hsu A, and Topol EJ. Use of antioxidant vitamins for the prevention of cardiovascular disease: meta-analysis of randomised trials. *Lancet* 361: 2017–2023, 2003.
- 151. Wang YY, Yang YX, Zhe H, He ZX, and Zhou SF. Bardoxolone methyl (CDDO-Me) as a therapeutic agent: an update on its pharmacokinetic and pharmacodynamic properties. *Drug Des Devel Ther* 8: 2075–2088, 2014.
- 152. Welsh JL, Wagner BA, van't Erve TJ, Zehr PS, Berg DJ, Halfdanarson TR, Yee NS, Bodeker KL, Du J, Roberts LJ, 2nd, Drisko J, Levine M, Buettner GR, and Cullen JJ. Pharmacological ascorbate with gemcitabine for the control of metastatic and node-positive pancreatic cancer (PACMAN): results from a phase I clinical trial. *Cancer Chemother Pharmacol* 71: 765–775, 2013.

- 153. Wendel A, Fausel M, Safayhi H, Tiegs G, and Otter R. A novel biologically active seleno-organic compound—II. Activity of PZ 51 in relation to glutathione peroxidase. *Biochem Pharmacol* 33: 3241–3245, 1984.
- 154. Wind S, Beuerlein K, Armitage ME, Taye A, Kumar AH, Janowitz D, Neff C, Shah AM, Wingler K, and Schmidt HH. Oxidative stress and endothelial dysfunction in aortas of aged spontaneously hypertensive rats by NOX1/2 is reversed by NADPH oxidase inhibition. *Hypertension* 56: 490–497, 2010.
- 155. Xia N, Daiber A, Habermeier A, Closs EI, Thum T, Spanier G, Lu Q, Oelze M, Torzewski M, Lackner KJ, Munzel T, Forstermann U, and Li H. Resveratrol reverses endothelial nitric-oxide synthase uncoupling in apolipoprotein E knockout mice. *J Pharmacol Exp Ther* 335: 149–154, 2010.
- 156. Yeom CH, Lee G, Park JH, Yu J, Park S, Yi SY, Lee HR, Hong YS, Yang J, and Lee S. High dose concentration administration of ascorbic acid inhibits tumor growth in BALB/C mice implanted with sarcoma 180 cancer cells via the restriction of angiogenesis. *J Transl Med* 7: 70, 2009.
- 157. Yusuf S, Dagenais G, Pogue J, Bosch J, and Sleight P. Vitamin E supplementation and cardiovascular events in highrisk patients. The Heart Outcomes Prevention Evaluation Study Investigators. *N Engl J Med* 342: 154–160, 2000.
- 158. Yusuf S, Sleight P, Pogue J, Bosch J, Davies R, and Dagenais G. Effects of an angiotensin-converting-enzyme inhibitor, ramipril, on cardiovascular events in high-risk patients. The Heart Outcomes Prevention Evaluation Study Investigators. N Engl J Med 342: 145–153, 2000.

E-mail: daiber@uni-mainz.de

Dr. Antonio Cuadrado Instituto de Investigaciones Biomédicas "Alberto Sols" UAM-CSIC C/Arturo Duperier 4 Madrid 28029 Spain

E-mail: antonio.cuadrado@uam.es

Date of first submission to ARS Central, May 23, 2015; date of final revised submission, June 26, 2015; date of acceptance, July 3, 2015.

#### **Abbreviations Used**

AKT = protein kinase B

ARE = antioxidant response element

 $BH_4 = tetrahydrobiopterin$ 

BO-653 = 2,3-dihydro-5-hydroxy-2,2-dipentyl-4,6-di-tert-butylbenzofuran

CDDO; CDDO-Im = 2-cyano-3,12-dioxooleana-1,9-dien-28-oic acid-imidazolide

CVD = cardiovascular disease

DMF = dimethyl fumarate

eNOS = endothelial nitric oxide synthase

ERK = extracellular signal-regulated kinases

 $gp91^{phox} = 91kDa$  membranous subunit of the phagocyte NADPH oxidase (Nox2)

GSH = glutathione

 $H_2O_2 = hydrogen peroxide$ 

JNK = c-Jun N-terminal kinases

KEAP1 = kelch-like ECH-associated protein 1

MAPK = mitogen-activated protein kinase

 $\label{eq:mirNA} \begin{aligned} & \text{mirNA} = \text{microRNA, a small noncoding RNA} \\ & & \text{molecule} \end{aligned}$ 

mitoTEMPO = (2-(2,2,6,6-Tetramethylpiperidin-1-oxyl-4-ylamino)-2-oxoethyl)triphenylphosphonium chloride monohydrate

MMF = monomethyl fumarate

MS = multiple sclerosis

NAC = N-acetylcysteine

NADPH = nicotinamide adenine dinucleotide phosphate

•NO = nitric oxide

NOS = nitric oxide synthase

NOX = nicotinamide adenine dinucleotide phosphate oxidases

(1, 2, 4 isoforms)

NRF2 = nuclear factor (erythroid-derived 2)like 2

NXY-059 = disufenton sodium, a synthetic antioxidant

 $O_2^{-\bullet}$  = superoxide anion

p38 = p38 mitogen-activated protein kinases

PGC1 $\alpha$  = peroxisome proliferator-activated receptor gamma coactivator  $1\alpha$ 

PTEN = phosphatase and tensin homolog

ROS = reactive oxygen species

SOD = superoxide dismutase